

LETTER TO THE EDITOR

Assessing the effect of obesity-related traits on multiple myeloma using a Mendelian randomisation approach

Blood Cancer Journal (2017) 7, e573; doi:10.1038/bcj.2017.48; published online 16 June 2017

Multiple myeloma (MM) accounts for around 15% of new cases and 20% of deaths amongst patients diagnosed with haematological malignancy. To date, few risk factors have been robustly confirmed for MM, these include increasing age, male sex, black race and a family history of MM.¹

High body mass index (BMI) has been reported to be associated with an increased risk of MM in several observational studies, though questions remain regarding the aetiological relevance, including the distribution of body fat.² In addition to being a potential risk factor for MM, some, but not all, studies have suggested an association between BMI and prognosis.^{1,3} A recent study has suggested the relationship between BMI and MM may be through reduced levels of plasma adiponectin, the inflammatory mediator secreted by adipocytes. The association was, however, confined to obese individuals providing an argument against a direct causal link.⁴ Findings such as these, alongside the conflicting results of previous studies into adiposity traits, highlight the limitations of observational studies. Importantly, such studies do not establish a causal relationship, as they cannot fully eliminate the influence of confounding factors. Moreover, in the context of prognostication, many studies have not explicitly addressed the issue of reverse causation.⁵

Mendelian randomisation (MR) provides an attractive alternative to the traditional epidemiological study for examining relationships between exposure and disease. MR makes use of allelic variants, which are randomly assigned during meiosis and are robustly associated with traits of interest, as instrumental variables (IVs) to infer whether associations between exposure and disease are causal. The use of these genetically defined IVs as proxies of modifiable exposure, avoids confounding by environmental factors, can be reflective of life-long exposure and circumvents reverse causality.

Genome-wide association studies (GWAS) have identified single-nucleotide polymorphisms (SNPs) at multiple independent loci significantly associated with BMI, childhood obesity (CHO) and plasma levels of adiponectin.^{6–8} Here we have sought to establish a causal association between adiposity traits and MM by performing a MR analysis using SNPs associated with body mass index (BMI), hip circumference adjusted for BMI (HipadjBMI), waist circumference adjusted for BMI (WCadjBMI), waist-to-hip ratio adjusted for BMI (WHRadjBMI), CHO and plasma adiponectin levels as IVs.

We constructed genetic risk scores (GRS) to investigate the relationship of adiposity and plasma levels of adiponectin with MM risk, using the data from five reported MM GWASs, comprising 6 839 cases and 22 221 controls.⁹ We performed two-stage MR analysis to assess the association between each adiposity trait and MM using summary statistics from the MM GWAS, and the published effect size of the adiposity trait. As per Burgess *et al.*,¹⁰ a fixed-effects model was used to calculate combined ratio estimates, β , for the effect of each trait on MM risk. Results are summarised in Table 1.

Following calculation of combined ratio estimates, β , in three of the five cohorts, a positive association was shown between one s.d. in BMI (kg/m^2) and MM risk, with the UK-GWAS series being

nominally significant (Figure 1a). Meta-analysis of the data from all five cohorts did not, however, provide evidence for a causal association (odds ratio (OR) = 1.17, 95% confidence intervals (CIs): 0.92–1.49, $P=0.19$). Similarly, we found no association between the other anthropometric traits, which report on central obesity and MM risk, specifically—HipadjBMI, WCadjBMI, WHRadjBMI, which had respective ORs of 0.77 (95% CI: 0.42–1.41), 0.62 (95% CI: 0.37–1.02) and 0.82 (95% CI: 0.57–1.19) (Figures 1b–d). We also found no support for a relationship between CHO and MM risk (OR = 0.98, 95% CI: 0.87–1.10; Figure 1e). We then evaluated the impact of plasma levels of adiponectin on the risk of developing MM, again observing no association (OR = 1.04, 95% CI: 0.64–1.72; Figure 1f).

Linkage of the survival data to genotypes on three of the series allowed the relationship between the aforementioned adiposity-related traits and patient outcome to be examined through MR. In meta-analyses of these data, no association between any of the traits and either overall survival (OS) or progression-free survival (PFS) was shown.

The results from our study contrast with some observational epidemiological studies, which have shown a positive association between adiposity and MM risk and mortality.^{1,11} A recent meta-analysis of studies found a modest, but significant association between BMI and risk in prospective cohorts.¹¹ Reported relative risks for MM associated with obesity from these studies had OR 95% CIs of 1.08–1.35. Over this range of effect, we had 16–97% study power to demonstrate an association. Hence, we cannot exclude the possibility that the null results we observed were simply a consequence of limited study power if the true effect is marginal.

Obesity is a well-established risk factor for a number of other solid cancers.¹² By inference, it is likely *a priori* that obesity will also increase MM risk. An elevated level of insulin-like growth factor 1, associated with chronic hyperinsulinemia, stimulates cell proliferation and inhibits apoptosis, therefore providing a biological basis for obesity having a generic effect on cancer risk.¹³ In the case of MM, the insulin receptor is overexpressed in MM cells and is increased throughout normal plasma cell differentiation.¹⁴ These observations would therefore imply a causative effect of

Table 1. Results of associations of multiple myeloma (MM) risk with adiposity traits using Mendelian randomisation

Trait	Odds ratio	95% CI	P-value
BMI	1.17	0.92;1.49	0.19
CHO	0.98	0.87;1.10	0.71
WHRadjBMI	0.82	0.57;1.19	0.29
WCadjBMI	0.62	0.37;1.02	0.06
HipadjBMI	0.77	0.42;1.41	0.39
Adiponectin ^a	1.04	0.64;1.72	0.86

Abbreviations: BMI, body mass index; CHO, childhood obesity; HipadjBMI, hip circumference adjusted for BMI; WCadjBMI, waist circumference adjusted for BMI; WHRadjBMI, waist-to-hip ratio adjusted for BMI. Odds ratio, 95% confidence intervals (CIs) and P -values from meta-analysis of all cohorts, which demonstrates no significant causative effect of traits on MM risk. ^aMeta-analysis for adiponectin was conducted with 3 cohorts as no SNPs were present in UK and USA data sets.

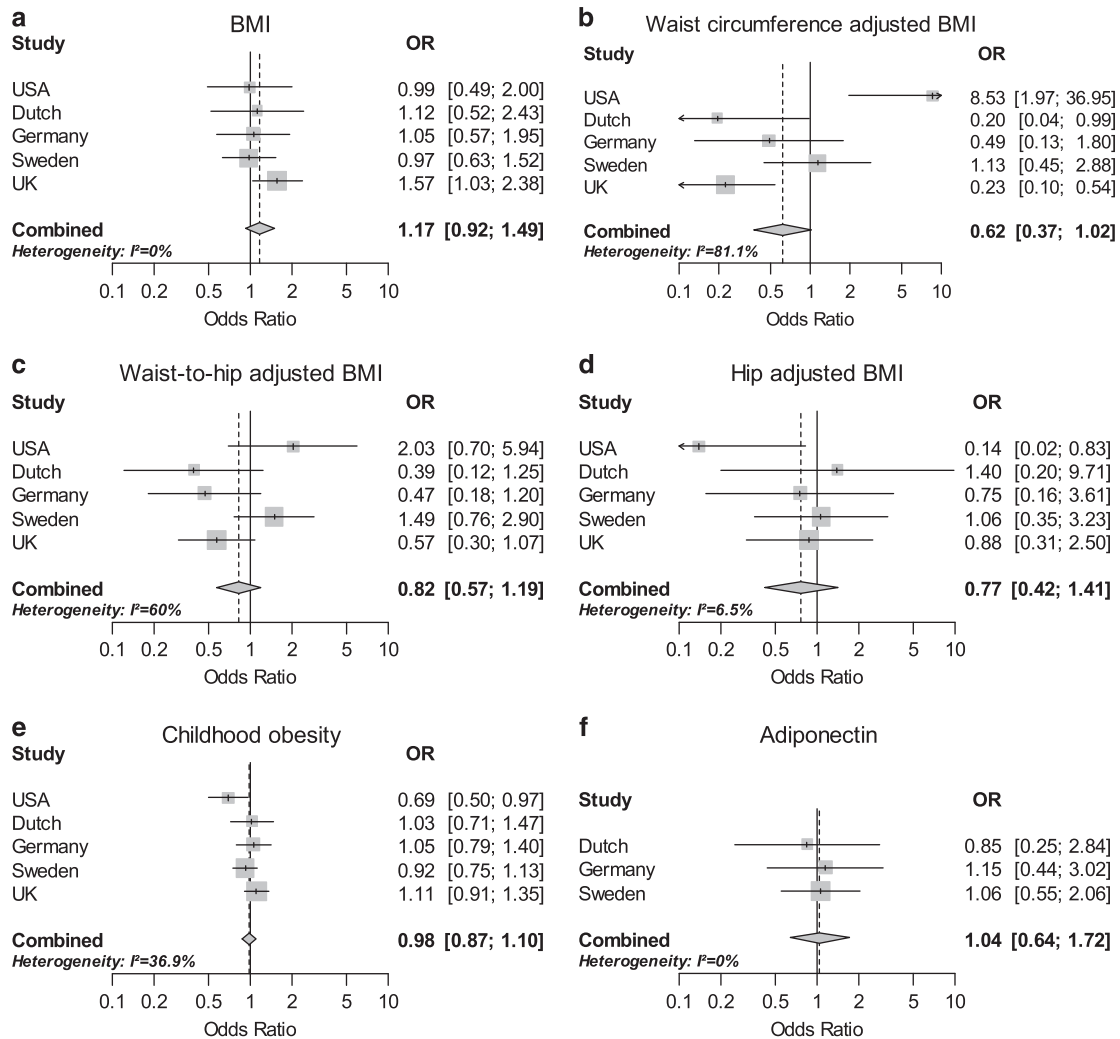


Figure 1. Meta-analysis odds ratios (OR) for multiple myeloma per unit increase in genetic risk score (s.d. trait) for each adiposity trait. Shaded boxes denote odds ratio for individual cohorts with areas proportional to the inverse variance weight of the estimate. Horizontal lines represent 95% confidence intervals (CIs) for individual cohorts. Shaded diamond represents summary ORs, computed under a fixed-effects models and diamond width gives 95% CIs. Solid vertical line represents null hypothesis (OR = 1) and dashed vertical line indicates OR from meta-analysis. (a) BMI. (b) Waist circumference adjusted for BMI. (c) Waist-to-hip ratio adjusted for BMI. (d) Hip circumference adjusted for BMI. (e) Childhood obesity. (f) Adiponectin levels.

obesity and ensuing hyperinsulinemia on plasma and myeloma cell growth; a corollary of this may be increased risk of MM and adverse patient outcome.

Our MR analysis does not suffer from the influence of recall bias and confounding that can affect observational studies.⁵ Nevertheless, a central assumption in MR is that the variants used as IVs are associated with the exposure being investigated. To ensure this was the case, we only used SNPs associated with adiposity-related traits at genome-wide significance from GWAS. Furthermore, we only used the data from individuals of European descent to limit bias from population stratification. An additional assumption is that the variants are associated with MM only through the exposure and are not confounded by pleiotropy. We assessed the impact of possible pleiotropism on MR estimates using both inverse variance weighted (IVW) and MR-Egger regression tests as per Bowden *et al.*¹⁵ Neither test provided evidence for pleiotropy, with respective P -values of 0.2 and 0.71 for BMI, 0.77 and 0.65 for CHO, 0.56 and 0.52 for HipadjBMI, 0.13 and 0.66 for WCadjBMI, 0.4 and 0.79 for WHRadjBMI, and 0.87 and 0.2 for adiponectin plasma levels. While we found no evidence that the SNPs violated this IV

assumption, this does not exclude confounding by as yet unidentified confounders.

In conclusion, high BMI is a plausible risk factor for MM; however, observational studies so far have provided varied and conflicting results, likely attributed to confounding and reverse causation. Our MR study, which uses IVs to avoid such confounding factors, provides no evidence of BMI or other tested adiposity traits, influencing MM risk or survival. To robustly establish a causal relationship through MR-based analyses, and thus avoid confounding, far larger datasets will be required than the one we have analysed. Such studies should be possible in the future with ongoing GWASs of MM currently being undertaken.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGEMENTS

In the United Kingdom, Myeloma UK and Bloodwise provided principal funding. Additional funding was provided by Cancer Research UK (C1298/A8362 supported by

the Bobby Moore Fund) and The Rosetrees Trust. This study made use of the genotyping data on the 1958 Birth Cohort generated by the Wellcome Trust Sanger Institute (<http://www.wtccc.org.uk>). We are grateful to all investigators who contributed to NSCCG and GELCAPS, from which controls in the replication were drawn. We also thank the staff of the CTRU University of Leeds and the NCRI Haematology Clinical Studies Group. The US GWAS was supported by a grant from the National Institutes of Health (P01CA055819). The German study was supported by the Dietmar-Hopp-Stiftung, Germany, the German Cancer Aid (110,131), the German Federal Ministry of Education and Research (CLIOMMICS 01ZX1309), the German Research Council (DFG; Project SI236/8-1, SI236/9-1, ER 155/6-1 and the DFG CRU 216) and the Multiple Myeloma Research Foundation. The patients were collected by the GMMG and DSMM studies. The German GWAS made use of the genotyping data from the population-based HNR study, which is supported by the Heinz Nixdorf Foundation (Germany). The genotyping of the Illumina HumanOmni-1 Quad BeadChips of the HNR subjects was financed by the German Center for Neurodegenerative Disorders (DZNE), Bonn, Germany. We are grateful to all investigators who contributed to the generation of this data set. The German replication controls were collected by Peter Bugert, Institute of Transfusion Medicine and Immunology, Medical Faculty Mannheim, Heidelberg University, German Red Cross Blood Service of Baden-Württemberg-Hessen, Mannheim, Germany. This work was supported by research grants from the Swedish Foundation for Strategic Research (KF10-0009), the Marianne and Marcus Wallenberg Foundation (2010.0112), the Knut and Alice Wallenberg Foundation (2012.0193), the Swedish Research Council (2012–1753), the Royal Swedish Academy of Science, ALF grants to the University and Regional Laboratories (Labmedicin Skåne), the Siv-Inger and Per-Erik Andersson Foundation, the Medical Faculty at Lund University and the Swedish Society of Medicine. We thank Jörgen Adolfsson, Tomas Axelsson, Anna Collin, Ildikó Frigyes, Patrik Magnusson, Bertil Johansson, Jan Westin and Helga Ögmundsdóttir for their assistance. We are indebted to the clinicians who contributed samples to Swedish, Norwegian and Danish biobanks. We are indebted to the patients and other individuals who participated in the project. This work was supported by Center for Translational Molecular Medicine (BioCHIP), a clinical research grant from the European Hematology Association, an EMCR Translational Research Grant, a BMBF grant from CLIOMMICS (01ZX1309A) and FP7 grant MSCNET (LSHC-Ct-2006-037602). We thank the staff of the HOVON, as well as patients and physicians at participating sites. In addition, we also thank Jasper Koenders, Michael Vermeulen, André Uitterlinden and Nathalie van der Velde for their assistance.

M Went^{1,2}, A Sud¹, PJ Law¹, DC Johnson², N Weinhold^{3,4}, A Försti^{5,6}, M van Duin⁷, JS Mitchell¹, B Chen⁵, R Kuiper⁷, OW Stephens³, U Bertsch^{4,8}, C Campo⁵, H Einsele⁹, WM Gregory¹⁰, M Henrion¹, J Hillengass⁴, P Hoffmann^{11,12}, GH Jackson¹³, O Lenive¹, J Nickel⁴, MM Nöthen^{11,14}, MI da Silva Filho⁵, H Thomsen⁵, BA Walker³, A Broyl⁷, FE Davies³, C Langer¹⁵, M Hansson^{16,17}, M Kaiser⁷, P Sonneveld⁷, H Goldschmidt^{4,8}, K Hemminki^{5,6}, B Nilsson^{16,18,19}, GJ Morgan³ and RS Houlston¹

¹Division of Genetics and Epidemiology, The Institute of Cancer Research, London, UK;
²Division of Molecular Pathology, The Institute of Cancer Research, London, UK;
³Myeloma Institute for Research and Therapy, University of Arkansas for Medical Sciences, Little Rock, AR, USA;
⁴Department of Internal Medicine V, University of Heidelberg, Heidelberg, Germany;
⁵Molecular Genetic Epidemiology, German Cancer Research Center, Heidelberg, Germany;
⁶Center for Primary Health Care Research, Lund University, Malmö, Sweden;
⁷Department of Hematology, Erasmus MC Cancer Institute, Rotterdam, The Netherlands;
⁸National Center for Tumor Diseases, Heidelberg, Germany;
⁹Department of Internal Medicine II, Division of Hematology and Medical Oncology, University Hospital Würzburg, Würzburg, Germany;
¹⁰Clinical Trials Research Unit, Leeds Institute of Clinical Trials Research, University of Leeds, Leeds, UK;
¹¹Institute of Human Genetics, University of Bonn, Bonn, Germany;
¹²Division of Medical Genetics, Department of Biomedicine, University of Basel, Basel, Switzerland;

¹³Royal Victoria Infirmary, Newcastle upon Tyne, Newcastle, UK;
¹⁴Department of Genomics, Life & Brain Center, University of Bonn, Bonn, Germany;
¹⁵Department of Internal Medicine III, University of Ulm, Ulm, Germany;
¹⁶Division of Hematology and Transfusion Medicine, Department of Laboratory Medicine, Lund University, Lund, Sweden;
¹⁷Hematology Clinic, Skåne University Hospital, Lund, Sweden;
¹⁸Clinical Immunology and Transfusion Medicine, Laboratory Medicine, Office of Medical Services, Lund, Sweden and
¹⁹Broad Institute, 7 Cambridge Center, Cambridge, MA, USA
 E-mail: richard.houlston@icr.ac.uk

REFERENCES

- Teras LR, Kitahara CM, Birmann BM, Hartge PA, Wang SS, Robien K *et al*. Body size and multiple myeloma mortality: a pooled analysis of 20 prospective studies. *Br J Haematol* 2014; **166**: 667–676.
- Carson KR, Bates ML, Tomasson MH. The skinny on obesity and plasma cell myeloma: a review of the literature. *Bone Marrow Transplant* 2014; **49**: 1009–1015.
- Beason TS, Chang SH, Sanfilippo KM, Luo S, Colditz GA, Vij R *et al*. Influence of body mass index on survival in veterans with multiple myeloma. *Oncologist* 2013; **18**: 1074–1079.
- Hofmann JN, Birmann BM, Teras LR, Pfeiffer RM, Wang Y, Albanes D *et al*. Low levels of circulating adiponectin are associated with multiple myeloma risk in overweight and obese individuals. *Cancer Res* 2016; **76**: 1935–1941.
- Berrigan D, Troiano RP, Graubard BI. BMI and mortality: the limits of epidemiological evidence. *Lancet* 2016; **388**: 734–736.
- Dastani Z, Hivert MF, Timpson N, Perry JR, Yuan X, Scott RA *et al*. Novel loci for adiponectin levels and their influence on type 2 diabetes and metabolic traits: a multi-ethnic meta-analysis of 45,891 individuals. *PLoS Genet* 2012; **8**: e1002607.
- Locke AE, Kahali B, Berndt SI, Justice AE, Pers TH, Day FR *et al*. Genetic studies of body mass index yield new insights for obesity biology. *Nature* 2015; **518**: 197–206.
- Shungin D, Winkler TW, Croteau-Chonka DC, Ferreira T, Locke AE, Magi R *et al*. New genetic loci link adipose and insulin biology to body fat distribution. *Nature* 2015; **518**: 187–196.
- Mitchell JS, Li N, Weinhold N, Forsti A, Ali M, van Duin M *et al*. Genome-wide association study identifies multiple susceptibility loci for multiple myeloma. *Nat Commun* 2016; **7**: 12050.
- Burgess S, Butterworth A, Thompson SG. Mendelian randomization analysis with multiple genetic variants using summarized data. *Genet Epidemiol* 2013; **37**: 658–665.
- Wallin A, Larsson SC. Body mass index and risk of multiple myeloma: a meta-analysis of prospective studies. *Eur J Cancer* 2011; **47**: 1606–1615.
- De Pergola G, Silvestris F. Obesity as a major risk factor for cancer. *J Obes* 2013; **2013**: 291546.
- Kumari N, Dwarakanath BS, Das A, Bhatt AN. Role of interleukin-6 in cancer progression and therapeutic resistance. *Tumour Biol* 2016; **37**: 11553–11572.
- Sprynski AC, Hose D, Kassambara A, Vincent L, Jourdan M, Rossi JF *et al*. Insulin is a potent myeloma cell growth factor through insulin/IGF-1 hybrid receptor activation. *Leukemia* 2010; **24**: 1940–1950.
- Bowden J, Davey Smith G, Burgess S. Mendelian randomization with invalid instruments: effect estimation and bias detection through Egger regression. *Int J Epidemiol* 2015; **44**: 512–525.



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>