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**Reproductive and population genetic characteristics of leading-edge and central populations of two temperate forest tree species and implications for range expansion**

**Running Title:** Genetics of leading-edge vs central tree populations

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26 **Abstract**

27 It is still a matter of debate how reproductive and genetic characteristics of range-edge  
28 populations differ from those central to its range, yet this is important for future changes in  
29 species' ranges. Here we use microsatellite markers to assess the genetic diversity,  
30 relatedness and clonal reproduction of two lime tree species, *Tilia cordata* and *T.*  
31 *platyphyllos*, from leading range-edge and central locations.

32 Clonal reproduction was limited in the populations studied, but leading range-edge  
33 populations contained more clones than populations sampled from lower latitudes. Although  
34 no inbreeding or lower genetic diversity was detected, leading range-edge populations have  
35 a higher average relatedness of individuals and a lower effective population size than those  
36 populations closer to the centre of the species' distribution. Trees further apart than 26m are  
37 unlikely to be clones and those further apart than 61m are not likely to be closely related.

38 The implications for forest managers are that although leading range-edge  
39 populations have a lower effective population size, they are also likely to be better adapted to  
40 northern climes. Because locally sourced trees appear to have sufficient genetic diversity  
41 and predominantly result from sexual reproduction they are likely to adapt to climate change  
42 and be suited for natural migration and a good source for assisted migration. This is a  
43 promising prospect for the potential of future natural or managed expansion and increased  
44 species abundance.

45

46 **Keywords:** Assisted migration; Climate change; Clonal reproduction; Leading range-edge;  
47 Microsatellites; Relatedness; Range shifts; *Tilia*

48

## 49        1. Introduction

50 Range-edge populations exist in less than optimum conditions, and are often smaller and  
51 more isolated than central populations. , The majority of the studies indeed show that range-  
52 edge populations are vulnerable to loss of genetic diversity and increased population  
53 differentiation through random genetic drift (e.g. Arnaud-Haond *et al.*, 2006; Eckert *et al.*,  
54 2008; Gapare & Aitken, 2005). However, a few phylogeographic studies have suggested that  
55 current genetic diversity has been influenced by historical range expansions and contractions  
56 leading to boundary populations in some species that contain a larger proportion of the  
57 species diversity compared to the central populations (Hewitt, 2004; Petit *et al.*, 2003). In  
58 addition, the demography and mode of reproduction in range-edge populations may also  
59 differ from central populations (Dorken & Eckert, 2001; Vucetich & Waite, 2003), potentially  
60 also affecting genetic diversity, population differentiation and the ability to adapt.

61        Asexual reproduction is a survival mechanism common in plants and is achieved  
62 through agamospermy or clonal (vegetative) reproduction e.g. root suckering, root collar  
63 growth, epicormic shoots, (Bond & Midgley, 2001; Ellstrand & Roose, 1987; Evans & Morris,  
64 2016; Widén *et al.*, 1994). Clonality can be seen as an adaptation to persist in adverse  
65 conditions (Macaya-Sanz *et al.*, 2016; Meloni *et al.*, 2013) and offers both advantages and  
66 disadvantages. For example, being clonal prolongs the lifespan of an individual which may  
67 lead to lower genetic diversity among adult stems (De Witte & Stöcklin, 2010). Also, clonal  
68 organisms suspend the exchange and recombination of genetic material, and thus fewer  
69 novel genotypes are generated. If sexual reproduction is limited to only a few individuals then  
70 asexual reproduction is advantageous as otherwise inbreeding and random allele loss may  
71 occur (Ennos, 2003), and genetic diversity is maintained (Balloux *et al.*, 2003). In addition,  
72 large clones may accumulate somatic mutations, potentially generating diversity and  
73 elevating heterozygosity (Balloux *et al.*, 2003; Halkett *et al.*, 2005). It is also possible  
74 selection plays a role, i.e. higher survival of more heterozygous genotypes. However,

75 adaptation, for example to a changing climate, is limited in largely clonally reproducing  
76 populations. On the contrary, sexual reproduction would produce an excess of seedlings for  
77 natural selection to act on. Therefore, the differences in relative amounts of sexual and  
78 asexual reproduction can change the genetic composition, diversity and adaptability of  
79 organisms.

80 In several plant species, strong clonal reproduction, low levels of genetic diversity and  
81 high population differentiation in range-edge populations, as compared to those at the centre  
82 of the species' range, have been reported. These studies concentrated often on shrubs and  
83 weeds (e.g. Beatty *et al.*, 2008; Dorken & Eckert, 2001), but some significant work has also  
84 been carried out on long-lived organisms, such as forest trees (e.g. Jankowska-Wroblewska  
85 *et al.*, 2017; Santos-del-Blanco *et al.*, 2013; Stoeckel *et al.*, 2006; Vaughan *et al.*, 2007).  
86 Although clonal spread certainly occurs in forest trees, these are generally not considered to  
87 be species where clonal propagation is the predominant form of reproduction.

88 Trees from the genus *Tilia* were important components of woodlands in Europe until a  
89 few centuries ago, and the woodlands where they currently exist are largely remnants of their  
90 original distribution (Pigott, 2012). As a survival strategy, in sub-optimum conditions (e.g.  
91 cooler temperatures resulting in shorter flowering period and/or fewer pollinating insects),  
92 members of the genus may reproduce asexually (Evans & Morris, 2016; Logan *et al.*, 2018).  
93 *Tilia*, may be considered a partially clonal reproducing species, but the extent of asexual  
94 reproduction and intra-population relatedness across their range is not known. Therefore,  
95 *Tilia* serves as an excellent forest tree model to investigate leading range-edge effects,  
96 adding valuable information to the existing literature (e.g. Myking, 2002; Siefert *et al.*, 2015).

97 Species within the genus *Tilia* tend to cope well with cyclic management regimes,  
98 such as coppicing (Buckley & Mills, 2015; Pigott, 2012). Coppicing encourages sprouting,  
99 and therewith prolongs the lifespan and (clonal) spread of a genotype. Clonal spread is also  
100 aided by land erosion, particularly on steep banks and cliffs and individual trees can spread  
101 through low hanging branches and fallen trunks (S. A. Logan *pers. obs.*). The ability to

102 regenerate through vegetative growth and the low rate of regeneration by seed at some  
103 locations, e.g. range-edge (attributed to the present climate), may explain current  
104 distributions (Pigott & Huntley, 1978). Earlier work has shown that populations in the UK  
105 clonally reproduce at some sites (Logan *et al.*, 2015; Mylett, 2015; Phuekvilai, 2014).  
106 Likewise, Novák *et al.* (2014) suggest that *T. sibirica* (Siberian lime) shows evidence of clonal  
107 growth, which has been confirmed by microsatellite genotyping (Logan *et al.*, 2018). Also,  
108 remnant stands of *T. americana* (American basswood) on islands off the coast of Georgia  
109 (USA) largely depend on vegetative growth (Evans & Morris, 2016).

110 Radoglou *et al.* (2009), and references cited therein, suggest that *T. cordata* in south-  
111 western Russia has a greater incidence of clonal reproduction than sexual reproduction,  
112 claiming that nearly all *Tilia* in the north-east of its European distribution originated from  
113 vegetative reproduction. They also claim that 90% of *the Tilia* trees in the Strict Reserve of  
114 the Białowieża National Park (BNP) sprout from root collars, and that this phenomenon  
115 extends to the range-edge populations in the UK, Finland, and Siberia. These statements,  
116 based on morphology, need genetic confirmation. In contrast, it has been noted that *T.*  
117 *cordata* regenerates freely from seedlings in the BNP (Pigott, 1975) and in other areas of  
118 Poland (Jaworski *et al.*, 2005). Clonal reproduction was certainly not as high as 90% in  
119 sampled UK populations (Logan *et al.*, 2015), and while *T. cordata* trees from western  
120 Siberia did show some clonal occurrence, lime trees from BNP in Poland appear not to be  
121 clonal but largely a result of sexual reproduction (Logan *et al.*, 2018).

122 Climate change is altering both the potential and actual distribution, of many animal  
123 and plant species (Gibson *et al.*, 2009; Rehm *et al.*, 2015; Walther *et al.*, 2002). Species may  
124 need to track their climate induced range shifts (Essl *et al.*, 2015) to higher latitudes, where it  
125 is colder - the leading edge, or to lower latitudes, where it is warmer - the trailing edge  
126 (Hampe & Petit, 2005; Hewitt, 2004). Here we concentrate on a comparison between leading  
127 edge (northerly) populations with those from regions that are central in the species'  
128 distribution. Due to the rapidity of climate change, the shift of ranges suitable for habitation

129 and sexual reproduction of the species may not be synchronized with the actual migration of  
130 organisms (Hoegh-Guldberg *et al.*, 2008; McLachlan, *et al.*, 2007; Seddon, 2010; Walther *et*  
131 *al.*, 2002). The migration rates of trees, and in particular of *Tilia*, in the past were much  
132 slower than migration rates needed now for some organisms to track potential ranges  
133 (McLachlan *et al.*, 2005; Myking, 2002), also due to human habitat use and geography.  
134 Therefore, in addition to natural migration, assisted migration, *i.e.* transfer of trees or planting  
135 of seeds, may be needed (Aitken & Whitlock, 2013; Seddon, 2010; Whittet *et al.*, 2016;  
136 Winder *et al.* 2011).

137         Trees from populations nearer to their central ecological/geographical area are  
138 unlikely to have the adaptation for optimal reproduction and survival at higher latitudes  
139 further north, *e.g.* differences in season and day length or winter and late frost resistance  
140 (Kreyling *et al.*, 2014; Rehm *et al.* 2015) . Therefore, genetically diverse populations with a  
141 diversity of microhabitats outside of the main central range might be key to the survival of  
142 some tree species as they are more likely to harbor pre-adaptation to environmental change  
143 (Hampe & Petit, 2005; Lobo *et al.*, 2018b). This is particularly important in species such as  
144 *Tilia*, where season length is crucial for sexual reproduction. Although there is a view that  
145 range-edge populations may not be optimally adapted to their habitat due to persistent gene  
146 flow from central populations (*e.g.* Mimura & Aitken, 2007), it is likely that if there is sexual  
147 reproduction in a population there will be ample seedlings upon which selection can act.  
148 Seed and pollen dispersal in *Tilia* species has not yet been assessed, but is expected to be  
149 limited compared to other tree species (Myking, 2002) because *Tilia* is insect pollinated and  
150 seeds are heavy and wind-dispersed. Therefore, persistent gene flow from central to range-  
151 edge population is unlikely to be the case in *Tilia*, in particular with (currently) fragmented  
152 populations. Moreover, there is evidence for adaptation to latitude as spring phenology  
153 adaptation has been recorded in *T. cordata* (Lobo *et al.*, 2018a).

154         Microsatellite markers indicated high levels of polymorphism in *T. cordata* and *T.*  
155 *platyphyllos* (Logan *et al.*, 2015; Phuekvilai & Wolff, 2013) and allowed identification of

156 clones. Here we investigate clonal architecture, relatedness and genetic diversity at leading  
157 range-edge populations in comparison and central populations, using two putative partially  
158 clonal *Tilia* tree species. In particular, this study investigates; [1] the level of clonality, [2] the  
159 within-population relatedness of individuals and  $F_{IS}$ , [3] genetic diversity and structure, and  
160 [4] the effective population size of populations. We expect that in regions with hampered  
161 sexual reproduction (northern range-edge), asexual reproduction must be more pronounced  
162 and that genetic diversity and effective population size will be lower, while population  
163 differentiation and relatedness will be higher than in central populations. The study will  
164 indicate whether range-edge populations of *Tilia* are potentially suitable for natural and  
165 assisted migration, aiding future management of these woodland species.

166

## 167 **2. Materials and Methods**

### 168 2.1 *Study sites and sample collection*

169 Two species of temperate, long-lived forest tree, *Tilia cordata* Mill. (small-leaved lime) and *T.*  
170 *platyphyllos* Scop. (large-leaved lime) were studied. These two species are mostly found in  
171 European parts of the northern hemisphere and their distribution is limited because of poor  
172 seed set in the short and cool summers at the northern range-edge (Pigott & Huntley, 1978).  
173 The distribution of *T. platyphyllos* extends north to Sweden and south to Greece and Turkey,  
174 while its east-west range extends from Wales to Poland, Slovakia and Romania (Fig 1). *Tilia*  
175 *cordata* has a wider distribution, from Finland and Norway in the north to Greece in the  
176 south, while its east-west range extends from Wales and Spain to Siberia, Russia. Both  
177 species now mainly exist in native or 'ancient' woodlands.

178 We define leading (northern) range-edge populations as those occurring at  $>52^{\circ}\text{N}$  for  
179 *T. platyphyllos* and  $>54^{\circ}\text{N}$  for *T. cordata* (Table 1, Fig 1). Populations sampled from  
180 locations between latitudes  $43^{\circ} - 52^{\circ}\text{N}$  for *T. platyphyllos* and  $43^{\circ} - 54^{\circ}\text{N}$  for *T. cordata* are  
181 considered central populations. UK samples were from Sites of Special Scientific Interest

182 and National Nature Reserves. Other collection sites were remnants of ancient woods and  
183 not planted, as there is no history of planting *Tilia* in these woodlands. No further details were  
184 available about the stands. Only adult trees were included in this study, i.e. trees with a  
185 circumference of 10 cm or more. Coordinates were recorded for as many individuals as  
186 possible using a Garmin hand-held GPS unit. We calculated geographic distances between  
187 trees in GenAEx v6.502 (Peakall & Smouse, 2006; 2012). Locations could not be  
188 systematically sampled with the same distances between trees, due to the terrain and the  
189 number and density of trees. Average distances between sampled trees are presented  
190 (Table 1). In two populations (SWWC, RUVA) only one coordinate for the population was  
191 available and hence mean distances between sampled trees could not be calculated. Also, in  
192 POBF locations of nine trees were not available.

193

#### 194 *2.2 DNA extraction and microsatellite genotyping*

195 Leaf samples were dried and stored at -20°C until required for genomic DNA extraction,  
196 using the CTAB method (Morgan-Richards & Wolff, 1999). A multiplex Polymerase Chain  
197 Reaction (PCR) procedure amplified 12 nuclear microsatellite regions (Tc6, Tc937, Tc920,  
198 Tc8, Tc943, Tc4, Tc927, Tc915, Tc963, Tc5, Tc951 and Tc7) following Phuekvilai and Wolff  
199 (2013). Microsatellites were genotyped using an ABI 3130XL Genetic Analyser, scored using  
200 Genemapper (Applied Biosystems) and binned manually.

201

#### 202 *2.3 Clonal structure*

203 Deviations from Hardy-Weinberg equilibrium (HWE) were tested in GENEPOP on the web  
204 v4.2 (Raymond & Rousset, 1995; Rousset, 2008) and the presence of null alleles was  
205 tested in MICRO-CHECKER v2.2.3 (Van Oosterhout *et al.*, 2004). The combination of  
206 all alleles across all 12 markers together is called a Multi Locus Genotype (MLG). Individuals  
207 sharing MLG are likely to be clones. However, mutation rates, in particular for microsatellites,



208 can be high enough for mutations to occur in old clones, making genotypes of members of a  
209 clone (ramets) differ from each other for one or more alleles. Although such cases were  
210 checked and corrected for genotyping errors this could have contributed to ramets of a single  
211 clone being similar but not identical. In such a case ramets might have a MLG that differs by  
212 one or few alleles from other ramets of the same clone. In the literature this is sometimes  
213 called a Multi Locus Lineage (MLL). Hereafter, when we use the term 'clone' instead of MLL.

214 It is essential to determine the threshold at which MLGs should be considered to be  
215 part of a clone (Aizawa *et al.*, 2017; Meloni *et al.*, 2013). Microsatellite mutation rates and an  
216 objective criterion are not available. We used GenoDive 2.0b (Meirmans & Van Tienderen,  
217 2004) to determine the optimal threshold for assigning samples to clones, by determining the  
218 allele difference represented by the smallest number of pairs, using the infinite allele model.  
219 For this a matrix of genetic distance between all pairs of trees was calculated. Two trees  
220 identical for all alleles are clearly clones. Those differing for a small number of alleles could  
221 have arisen due to somatic mutation or due to genotyping errors. It is important to distinguish  
222 those that have clearly a different MLG (i.e. at multiple loci) from those that could erroneously  
223 be considered different. The threshold indicates the maximum distance (alleles different)  
224 allowed between two trees for them to still be considered clone mates. The limit to be set is  
225 reached by constructing a histogram of the frequency of the allelic differences. Those to the  
226 left of the valley (defined as the number of alleles different with the lowest frequency) will be  
227 considered clone members and those to the right of the valley will be considered to be  
228 different. This analysis had strong power and showed that trees with a difference of  
229 maximally three alleles would be assigned to the same clone (Fig A1). Clonal spread was  
230 quantified by averaging the distance between trees belonging to the same clone.

231 Once the clones within each population were determined we used GenClone  
232 (Arnaud-Haond & Belkhir, 2007) to provide an unbiased (*i.e.* not affected by genet size)  
233 estimate of genotypic richness  $R = (G-1)/(N-1)$ , where N is the number of samples and G the  
234 number of genotypes.  $R$  is a modified measure of proportion distinguishable genotypes ( $Pd$ ,

235 Ellstrand & Roose, 1987). The value will be '0' when stands consist of a single clone and '1'  
236 when all sampled trees are separate genets (Dorken & Eckert, 2001). To describe clonal  
237 heterogeneity, an adapted estimate of the Simpson's complement index (*i.e.*  $D^*$ ),  
238 independent of sample size (Pielou, 1969), was calculated. The Simpson index ( $D$ )  
239 represents the probability that two randomly sampled plants belong to the same species  
240 (Simpson, 1949). The Simpson's complement index ( $1-D$ ) of diversity is commonly reported  
241 in clonal studies as  $D^*$  (Arnaud-Haond *et al.*, 2007) and ranges from '0' when all trees within  
242 a population are a single clone to '1' when they are all unique. Additionally, the statistics  $P_{gen}$ ,  
243 the probability of trees having the same genotype (*i.e.* are part of same clone) by chance  
244 following the methods of Parks and Werth (1993) and  $P_{sex}$ , the probability that a clonal  
245 genotype originates from sexual reproduction at the first reencounter, were estimated.  $P_{gen}$   
246 assumes populations are in HWE. An adjusted measure  $P_{gen}(f)$ , taking into account HWE  
247 departure can be estimated, providing a more conservative estimate of  $P_{sex}$  (Arnaud-Haond  
248 *et al.*, 2007). These last two estimates indicate the power of the markers to detect clones.

249

#### 250 2.4 Genetic diversity and differentiation

251 In the population genetic analyses we used single representatives of each clone, *i.e.* at genet  
252 level, reflecting genetic characteristics comparable between populations. Genetic diversity,  
253 calculated as the expected heterozygosity ( $H_e$ ), and the fixation index  $F_{IS}$  were calculated in  
254 GenAlEx 6.502. When testing the difference in  $F_{IS}$  between non-clonal and clonal genotypes  
255 the fixation index  $F_{IS}$  was calculated by hand as  $F = (H_e - H_o) / H_e$ , with  $H_e$  being the expected  
256 heterozygosity over all clonal genotypes in that population. AMOVA analyses were  
257 performed to quantify population differentiation of the leading range-edge and of central  
258 populations of each species in GenoDive 2.0b using the Weir and Cockerham (1984)  
259 method. In addition, GenoDive supplied  $G$ -statistics  $G_{ST}$  (Nei, 1987), adjusted  $G'_{ST}$   
260 (correcting for the bias due to small numbers of populations) as well as Jost's  $D_{est}$  (Jost,  
261 2008), which is independent of  $H_e$ . It also supplied standard deviations through jack-knifing

262 over loci and 95% confidence intervals (CI) through bootstrapping over loci. These statistics  
263 were tested in GenoDive (Compare Groups) for significance of difference between leading  
264 range-edge and central populations, using 9999 permutations. We tested Isolation by  
265 Distance (IBD) to determine if genetic distance between populations was correlated to  
266 geographic distance by performing a Mantel Test in GenAlEx.

267

## 268 *2.5 Relatedness and effective population size*

269 Relatedness and effective population size were estimated at both genet (with single  
270 representative per clone) and ramet level (including all samples), where ramets reflect the  
271 actual standing crop. ML-Relate (Kalinowski *et al.*, 2006) uses simulations to determine the  
272 most likely relationship (unrelated, half sib, full sib or parent-offspring) and compares this  
273 with the likelihood of other relationships for the same pair of trees. We used 100,000  
274 randomisations and counted for each population the number of pairs that have the highest  
275 probability of being full sib (50% related), parent-offspring (50% related) and half sibs (25%  
276 related). We calculated the number of first degree related pairs as well as the sum of first and  
277 second degree related pairs (50% relatedness and 50%+25% relatedness, respectively) as a  
278 proportion of the maximum number of pairs within a population ( $n * (n-1)/2$ , with  $n$  being the  
279 number of genotypes or ramets). To obtain a quantitative measure of relatedness we  
280 averaged the maximum likelihood estimates of relatedness ( $r$ , ML-Relate) for all pairs of  
281 trees in a population. The distance for each pair of genets within a population with a 50%  
282 relatedness was averaged to indicate the distance at which two trees can be closely related.

283 To detect small scale population structure, *i.e.* whether trees that were growing near  
284 to one another within a population were genetically more closely related than those at larger  
285 distances, the autocorrelation coefficient was calculated in GenAlEx. We tested whether the  
286 autocorrelation  $r$  as a function of the pairwise distance between trees is significantly different  
287 from zero and determined the 95% confidence intervals and error bars using 9999  
288 permutations and 9999 bootstraps. In each case we used eight even pairwise distance

289 classes, with sufficient pairwise comparisons per distance class (maximum sample distance  
290 in a population divided by eight).

291 We estimated the contemporary (or recent) effective population size ( $N_e$ ) using the  
292 molecular co-ancestry method of Nomura (2008) and the Linkage Disequilibrium (LD)  
293 method as implemented in NeEstimator V2.01 (Do *et al*, 2014). The LD method is based on  
294 Waples and Do (2008) and includes the Waples (2006) bias correction, which corrects the  
295 bias that could be introduced when sample size is lower than the actual effective population  
296 size. The 95% CIs are based on Jack-knifing over loci, and in the LD method only alleles with  
297 a frequency  $> 0.05$  were used.

298

## 299 2.6 *Contrasting groups of populations*

300 For testing the fixed-effect categorical variables of region, species and their interaction on  
301 clonal reproduction, genetic diversity and relatedness estimates we used averages per  
302 population. For most tests we used the average distance between trees (Av Dist) as a  
303 covariate to ensure that a difference in a measure was not biased by differences of within  
304 population sampling distances between the contrasting groups of populations. For the two  
305 populations without individual coordinates we replaced the missing Av Dist with the average  
306 for the group. The sample size (number of populations) was 28 for all tests. Three types of  
307 statistical analyses were carried out in R (R Core Development Team, 2016). For variables  
308 bounded between 0 and 1 ( $H_e$ ,  $r$ : ML-Relate, 1<sup>st</sup> and 2<sup>nd</sup> degree relatedness,  $D^*$  values) beta  
309 regression was performed (Ferrari & Cribari-Neto, 2004; betareg R package: Cribari-Neto &  
310 Zeileis, 2010). For variables from 0 to infinity ( $N_e$  values)  $\log(n+1)$  transformation of the  
311 values was used, with infinity set to 999. An infinite  $N_e$  estimate is likely caused by an  
312 actually large number, but the data do not allow an exact estimate (19 out of 60 estimates).  
313 The  $N_e$  data were analysed with a GLM model with Gaussian (normal) errors. For variables  
314 bounded by -1 and +1 ( $F_{IS}$ ) we used a logistic quantile regression (Bottai, *et al.*, 2010; lqr R  
315 package Galarza, *et al.*, 2016), and we could not use Av Dist as a covariate in this test.

316 Genetic diversity ( $H_e$ ) at the genet level were compared using a Welch's (unequal variance)  $t$ -  
317 test in R.

318 In addition, differences between range-edge and central populations for the degree of  
319 clonal reproduction ( $D^*$ ),  $H_e$ ,  $F_{IS}$  and population differentiation, were tested using the  
320 procedure 'compare groups' in GenoDive 2.0b using 9999 permutations with randomisation  
321 of populations over the groups. The difference in  $H_e$  and  $F_{IS}$  between non-clonal and clonal  
322 genotypes in populations was tested using a paired  $t$ -test in R.

323

### 324 **3. Results**

325 Genetic diversity was medium to high in both species for all populations. In *T. platyphyllos*  
326 the average number of alleles for the 12 microsatellite loci across all samples was 19.7, with  
327 a range of 8-27 across loci. Within *T. platyphyllos* populations this was an average of 7.6  
328 alleles per locus (range 3.1-9.8). For *T. cordata* the average number of alleles for the 12  
329 microsatellite loci across all samples was 13.7, with a range of 2-35 across loci. Within *T.*  
330 *cordata* populations this was an average of 5.8 alleles per locus (range 1.1-14.3). Deviations  
331 from HWE were observed at some loci, but were not consistent across populations. No null  
332 alleles were detected in the loci included in the analyses.

333

#### 334 *3.1 Clone identification*

335 In total 229 *T. platyphyllos* and 376 *T. cordata* trees were genotyped using microsatellite  
336 markers and clones were identified. Using a difference of three alleles as a maximum  
337 threshold value, ten trees with a unique MLG were deemed to be part of a clone. In all cases  
338 it produced clone members located closely together (average 5m for *T. platyphyllos* and 22m  
339 for *T. cordata*). The genotypes of members of a clone that did not have the exact same  
340 genotype, but differed for fewer alleles than the threshold value, were compared. Locus  
341 Tc963 was the most frequently deviating locus between clone-members, and therefore

342 seemed to accumulate mutations for members of a single clone (10 out of 17 presumed  
343 mutations). A total of 116 trees were not unique genotypes, *i.e.* belonged to a clone (19% of  
344 samples). Out of the total of 531 genotypes analysed, 489 were unique and 42 were  
345 represented by more than one tree (7.9%). Eight of the 12 populations of *T. platyphyllos* and  
346 six of the 16 *T. cordata* populations showed multiple adult trees belonging to a clone. No  
347 clones were found between locations. None of the clones were dominating a population, *i.e.*  
348 most clones (86%) were presented by two or three trees each (Table A2).

349         The number of loci screened was ample to identify MLG according to the resampling  
350 procedure in GenClone (data not presented, Arnaud-Haond et al., 2006). The probability of  
351 genotypes being identical by chance in a randomly mating population ( $P_{gen}$ ) as well as the  
352 probability that identical genotypes arose through sexual reproduction ( $P_{sex}$ ) were  $<0.001$  at  
353 all locations (Table S1), confirming that repeated genotypes were not due to chance and are  
354 in fact clones. Genotypic richness ( $R$ ) was high and rather variable across populations,  
355 ranging from 0.278 to 1.00 (Table A1). Clonal heterogeneity or Simpson's index ( $D^*$ ) was  
356 high in most populations, ranging from 0.743 to 1 (Table A1), suggesting a high probability  
357 that two randomly chosen trees were genetically different and a low rate of clonal  
358 reproduction.

359         The heterogeneity in clonal reproduction across populations was largely explained by  
360 'region', with range-edge populations having higher clonal reproduction (for  $R$ :  $P < 0.0005$  and  
361 for  $D^*$ :  $P < 0.0005$ ) (Table 2). The average distance between trees, used as covariate in the  
362 analysis, was not significant for both measurements ( $P = 0.053$  and  $0.099$ , respectively).  
363 Species and region x species interaction effects were not significant ( $P > 0.05$ ). Distances  
364 between trees belonging to a single clone was variable, on average 16.8m and top quintile  
365 26m, albeit smaller in *T. platyphyllos* than in *T. cordata* (average 5.7 and 19.3m, respectively,  
366 Fig. 2a and Table A3).

367

368 *3.2 Genetic diversity and fixation index*

369 Genetic diversity is represented by the expected heterozygosity ( $H_e$ ) and calculated at the  
370 genet level (Table A4).  $H_e$  was significantly higher in *T. platyphyllos* than in *T. cordata* ( $P$   
371  $<0.0005$ , Table 3). There was no effect of region, Av Dist or interaction on genetic diversity.  
372 Other measures of genetic diversity, such as effective number of alleles per locus, led to the  
373 same conclusions and are not presented here. Values of  $F_{IS}$  (fixation index) were variable  
374 across populations and reflect the outcrossing breeding system, ranging from -0.284 to  
375 0.105. However, there were no significant effects of species or region (species  $P = 0.729$ ,  
376 region  $P = 0.545$ , Table 3, Table A4).

377 To test whether large clones have accumulated mutations and have the potential to  
378 contribute to the maintenance of genetic diversity we contrasted genotypes that consist of  
379 single trees with those consisting of more than one tree (*i.e.* non-clonal vs clonal). This was  
380 done in the six *T. cordata* populations that had  $>2$  genotypes of both types. In all populations  
381 the observed heterozygosity ( $H_o$ ) was higher and the fixation index ( $F_{IS}$ ) lower in clonal than  
382 in non-clonal genotypes (Table 4). The difference in  $H_o$  is significant, both using a paired  $t$ -  
383 test ( $t = -3.73$ ,  $P = 0.014$ ) and using permutations in comparisons of groups in GenoDive ( $P =$   
384  $0.013$ ). The difference in  $F_{IS}$  between clonal and non-clonal genotypes is significant in a  
385 paired  $t$ -test ( $t = 3.60$ ,  $P = 0.015$ ), but is not significant using group comparison in GenoDive  
386 ( $P = 0.113$ ).

387

### 388 3.3 Population differentiation: contrast range-edge to central populations

389  $F_{ST}$  values were slightly higher in *T. platyphyllos* as a whole than in *T. cordata*, with 9% and  
390 7% of the variation between populations ( $F_{ST}$  0.090 and 0.070 and  $F'_{ST}$  0.369 and 0.180,  
391 respectively). Within each species the differentiation between populations was higher in the  
392 range-edge group than in the central group (Table 5 and 6).  $G_{ST}$ ,  $G'_{ST}$  and  $D_{est}$  were  
393 significantly higher for range-edge than central populations in *T. platyphyllos* ( $P$ -values  
394 0.014, 0.013 and 0.013, respectively), while this was significant for  $G'_{ST}$  and  $D_{est}$  in *T. cordata*  
395 ( $P$ -values 0.050, 0.040 and 0.003, respectively. The 95% confidence interval (CI) of  $F_{ST}$  for

396 range-edge and for central was not overlapping for *T. platyphyllos*, with 11.2% and 5.5% of  
397 variation between range-edge and central populations, respectively, while in *T. cordata* this is  
398 9.1% and 5.6%, respectively, with a small overlap of the 95% CI. A Mantel Test revealed  
399 slight positive correlation between genetic and geographic distances within all four groups: *T.*  
400 *platyphyllos* range-edge ( $R^2 = 0.208$   $P < 0.000$ ) and central ( $R^2 = 0.0811$   $P < 0.000$ ); *T.*  
401 *cordata* range-edge ( $R^2 = 0.0571$   $P < 0.000$ ) and central ( $R^2 = 0.0101$   $P = 0.001$ ).

402

### 403 3.4 Relatedness

404 The range-edge populations had a higher relatedness than populations from central regions,  
405 expressed as  $r$  (ML-Relate), but only at the ramet level ( $P < 0.0005$ ), and the Av Dist effect  
406 was also significant at the ramet level ( $P = 0.040$ ) (Table 7 and 8).  $R$  (ML-Relate) was not  
407 significantly different between the species ( $P = 0.079$  genotype level and  $P = 0.279$  ramet  
408 level).

409 To test the reliability of indicating the most likely relationships in ML-Relate we  
410 combined trees from different locations, expecting no relationships. We found some half-sib  
411 relationships across locations, albeit in much smaller numbers than within locations, and no  
412 parent-offspring or full sib relationships between locations. The fraction of first degree related  
413 pairs ranged from 0 to 0.056 at genet level and 0 to 0.038 at ramet level, while the fraction of  
414 first plus second degree relatedness ranged from 0 to 0.097 at genet level and 0.018 to  
415 0.086 at ramet level across populations (Table A4 and A5). Contrasting range-edge and  
416 central populations showed more related pairs in range-edge populations, only for the  
417 proportion of first degree relationships ( $P < 0.003$ ,  $P < 0.045$ , at genotype and ramet,  
418 respectively). There was no significant difference between species in fraction of first or first  
419 plus second degree relatedness. The co-variate Av Dist and the range x species interaction  
420 effects were not significant either. The average distance of parent-offspring and full sib pairs



421 was 52m across populations, with top quintile 61m (average 79m for *T. platyphyllos*, 34.4m  
422 for *T. cordata*, Figure 2b, Table A3).

423

### 424 3.5 *Small scale population structure*

425 The correlation of the autocorrelation coefficient  $r$  and geographic distance was tested at the  
426 genotype level so that the results were not biased due to ramets of the same clone being  
427 close together and having a relatedness of 1. Sixty percent of the populations (12 out of 20)  
428 for which we had individual tree locations and enough clones showed a positive correlation of  
429  $r$  with geographic distance in the first distance class (average 40m). Those populations that  
430 were collected with large distances between trees did not show autocorrelation in their first  
431 distance class.

432

### 433 3.6 *Effective population size*

434 Although the estimates for effective population size  $N_e$  differ between the estimation  
435 methods, coalescent and LD, and 95% CI are large, the values are likely to be comparable  
436 within this study. There was an effect of species for three of the four estimates ( $P = 0.048$   
437 and  $P = 0.042$ , at the ramet level and  $P = 0.009$  and  $P = 0.078$  at the genet level, for  $N_e$   
438 coalescent and LD, respectively). In all cases the  $N_e$  of *T. platyphyllos* was smaller than of *T.*  
439 *cordata*. Also, range-edge populations had lower  $N_e$  values than central populations ( $P =$   
440  $0.005$  and  $P < 0.001$  at the ramet level and  $P = 0.049$  and  $P = 0.031$  at the genotype level for  
441  $N_e$  coalescent and LD, respectively, Tables A2 and A4). The Av Dist or region x species  
442 interaction were not significant for any  $N_e$  estimate.

443

## 444 **4. Discussion**

445 Our study shows that leading range-edge populations of two *Tilia* species have more clonal  
446 reproduction than populations from central areas of their distribution. From  $F$ -statistics we

447 can conclude that range-edge populations are not likely to be affected by gene flow from  
448 other populations. This is largely concordant with several previous studies showing that  
449 range-edge populations, as compared to central range populations, have higher population  
450 differentiation ( $F_{ST}$ ) because of fragmentation and lower long distance gene flow, potentially  
451 enabling adaptation to current local conditions (Arnaud-Haond *et al.*, 2006; Eckert *et al.*,  
452 2008) or due to repeated extinction-founder effects. However, we did not find a significantly  
453 lower genetic diversity in range-edge populations, as some studies have indicated for other  
454 species (Arnaud-Haond *et al.*, 2006). Instead, *Tilia* range-edge populations have medium to  
455 high neutral genetic diversity (comparable to other trees species). However, provenance  
456 trials would be required to confirm that these range-edge populations also contain sufficient  
457 adaptive variation. With a warming climate, gene flow among populations may promote  
458 adaptation to novel conditions at northern latitudes.

459

#### 460 4.1 Clonality in *Tilia*

461 Clonal occurrence varies greatly across plant species (Ellstrand & Roose, 1987) and  
462 populations (Dorken & Eckert, 2001). Genotypic richness ( $R$ ) quantifies sexual versus  
463 asexual reproduction (Silvertown, 2008) and can be broadly compared across taxa. Clonality  
464 in *Tilia* at both the range-edge and central populations was generally low compared to other  
465 forest trees *e.g.* *Populus* species, *Prunus avium*, *P. ssiori*, *Ulmus minor*, and *Sorbus*  
466 *torminalis* (Fuentes-Utrilla *et al.*, 2014; Mock *et al.*, 2008; Nagamitsu *et al.*, 2004; Rasmussen  
467 & Kollmann, 2008; Santos-del-Blanco *et al.*, 2013; Stoeckel *et al.*, 2006; Vaughan *et al.*,  
468 2007). This is significant in that even in less optimal conditions, *Tilia* may still mostly  
469 reproduce sexually. Therefore, clonality was lower than expected from existing literature,  
470 which was based on observations and morphology (*e.g.* Radoglou *et al.*, 2009).

471 As expected, we found more clonal reproduction in range-edge than in central  
472 populations. High clonality was expected at Scandinavian, Russian and northerly UK sites  
473 due to range-edge effects and limited sexual reproduction (Pigott, 1981; Pigott & Huntley,

474 1981; Radoglou *et al.*, 2009), while clonality at central range sites, was expected to be lower  
475 due to greater sexual regeneration and a greater viability of seed (Pigott, 1981). Various  
476 additional factors can be involved in this. For example, *T. platyphyllos* trees at UK sites were  
477 large and old and with age comes more opportunity for clonal reproduction. In some  
478 populations there is also signs of past coppicing or self-coppicing due to steep terrain (*e.g.*  
479 UKBB). Other mechanisms, such as freezing/thawing, especially at northerly locations, or  
480 grazing can also have played a role in the tendency for sprouting (Morris *et al.*, 2014; Sjölund  
481 & Jump, 2014; Wilmking *et al.*, 2017).

482           Production of fertile seeds and regeneration seems to be a relatively recent  
483 phenomenon in UK *T. platyphyllos* (Pigott, 2000; 2012). Although references in Pigott (1975)  
484 state that fertile fruit are produced in Finland during very warm summers, our study suggests  
485 that asexual reproduction is important for maintaining *Tilia* in Finland where it competes with  
486 *Picea abies* (Pigott, 2012). Irregular sexual reproduction (of randomly mating individuals)  
487 intermittent to clonal reproduction permits the preservation of diversity and that diversity  
488 reflects the last time sexual reproduction occurred. Ennos (2003), reported that aspen  
489 showed high clonal occurrence and limited flowering but had similar genetic diversity to other  
490 outcrossing woody plants and attributes this survival to asexual reproduction of a previously  
491 random mating population which flowered regularly when conditions were more suitable. So  
492 essentially, genetic diversity was 'frozen in time' (Ennos, 2003).

493           We expected that other northern-most populations may have a similar ecology and  
494 demography to *Tilia* in Finland. However, we found no clones in the Norwegian population,  
495 but this is likely because samples were collected with large distances between trees (Table  
496 1) and so additional sampling from a smaller area is needed for a better understanding of the  
497 reproductive strategy in Norway. Climatic influences, topography and past management may  
498 also play a role in the clonal development of *Tilia* in Russia. Similar to *T. cordata* in our study,  
499 clonal occurrence has also been observed in *T. sibirica* (Siberian lime), further east in  
500 southern Siberia (Logan *et al.*, 2018; Novák *et al.*, 2014). Although, unlike many of the *Tilia*

501 populations sampled in this present study, *T. sibirica* showed low levels of genetic diversity  
502 (Mean  $H_e = 0.318$ ) and high levels of clonality (Mean  $R = 0.601$ ). Logan *et al.* (2018)  
503 attributed this to population fragmentation and difficulties for regeneration of seedlings due to  
504 competition from tall forbs, except in recent canopy gaps where *Tilia* seedlings may emerge  
505 (Novák *et al.*, 2014).

506 The BNP in Poland has had little human management (Miścicki, 2012). A recent  
507 study found no clones in the adult trees sampled from three plots within the Special Reserve  
508 (Logan *et al.*, 2018), and repeating the analyses here no clones were found. This contradicts  
509 Radoglou *et al.* (2009) who reported that *T. cordata* populations in Poland are highly clonal.  
510 In contrast rather, there were young trees present in the BNP (S. A. Logan, pers. obs.),  
511 aggregated in tight clusters, probably due to establishment of sexually produced seedlings in  
512 response to the creation of a recent gap in the canopy (Bobiec, 2007). The young trees from  
513 the BNP were not clones of any of the adult trees (Logan, 2016), confirming observations by  
514 Pigott (1975) and in other forests of southern Poland by Jaworski *et al.* (2005) that sexual  
515 reproduction is taking place there.

516 The concept that somatic mutations (or genotyping errors) contribute to a clone being  
517 composed of similar MLGs is accepted in other studies (e.g. James & McDougall, 2014). In  
518 an extreme case in *Robinia pseudoacacia* a 'hypervariable' locus was detected (Lian *et al.*,  
519 2004). In *Tilia*, the locus that seemed most mutable (Tc963) also has the largest number of  
520 alleles, namely 41. In addition, our finding that genets that are clonal had a higher observed  
521 heterozygosity than non-clonal genets (Table 4) follows expectations that large and long  
522 lived clones accumulate somatic mutations (Balloux *et al.*, 2003) and therewith potentially  
523 contribute to maintenance and generation of genetic diversity. It would be worth repeating  
524 this analysis with populations that have larger sample sizes. However, it may also be that  
525 trees with higher heterozygosity live longer and/or are more clonal than trees with lower  
526 heterozygosity (e.g. Vrankx *et al.*, 2014).

527 While clonal reproduction is an important survival tool for *Tilia* in several parts of its  
528 European range, sexual reproduction is still the most important mode of reproduction. This is  
529 particularly advantageous for range-edge populations: as the summers become warmer  
530 more viable seed will be produced and the exchange and recombination of genetic material  
531 will produce genotypes to allow for adaptation to warmer climates.

532

#### 533 4.2 Genetic diversity and differentiation

534 Genetic diversity ( $H_e$ ) in both *Tilia* species was similar to other partially clonal trees (Santos-  
535 del-Blanco *et al.*, 2013; Stoeckel *et al.*, 2006; Vaughan *et al.*, 2007). Range-edge populations  
536 do not have a significantly lower genetic diversity than those at central locations. This may be  
537 because range-edge populations are relicts of a diverse migration front that, through clonal  
538 reproduction, have maintained diversity or even increased genetic diversity through somatic  
539 mutations (Balloux *et al.*, 2003; Silvertown, 2008).  $H_e$  was higher in *T. platyphyllos* than in *T.*  
540 *cordata*, which could potentially be caused by ascertainment bias since the markers used  
541 were derived from *T. platyphyllos*. However, preliminary evidence using markers derived  
542 from *T. cordata* (Mylett, 2015) also revealed lower genetic diversity in *T. cordata* than in *T.*  
543 *platyphyllos* (Stephenson, Logan & Wolff *unpubl.*).

544 The near zero fixation index ( $F_{IS}$ ) reflects the outcrossing breeding system of the two  
545 species. There was no significant difference in  $F_{IS}$  values between range-edge and central  
546 populations, similar to *Acer* (Chybicki *et al.*, 2014). Our low  $F_{IS}$  values are in contrast to  
547 Ennos (2003), where he reported high  $F_{IS}$  values in clonal Aspen, likely due to a shortage of  
548 flowering mates. While we have no knowledge of the flowering status of the populations in  
549 our study, the  $F_{IS}$  values we report suggest that this is not the case in *Tilia*. The overall  
550 slightly negative  $F_{IS}$  values in the species and the slightly (but not significantly) lower  $F_{IS}$  in  
551 clonal range-edge populations may well be explained by somatic mutations (Balloux *et al.*,  
552 2003; Meloni *et al.*, 2013).

553            Assuming ideal populations the overall  $F_{ST}$  values seem low compared to some other  
554 studies and indicate migration rates ( $Nm$  values) that cannot be ignored. However, taking  
555 into account the long generation time and rare sexual reproduction, migration will have a  
556 small effect in these *Tilia* species. Similar to other studies (*e.g.* Arnaud-Haond *et al.*, 2006;  
557 Chybicki *et al.*, 2014) we confirmed here that range-edge populations show higher  
558 divergence from each other than central populations. It is likely that random genetic drift due  
559 to smaller effective population size and higher fragmentation is causing this. This may  
560 promote adaptation to current local conditions making them important for range expansion  
561 (Arnaud-Haond *et al.*, 2006). However, the difference in  $F_{ST}$  between range-edge and central  
562 populations could be an artefact, namely because range-edge populations are  
563 geographically further apart from each other than central populations. The weak but positive  
564 relationship between genetic and geographic distance (see section 3.3) hints at this.

565

#### 566 *4.3 Relatedness within populations and effective population size*

567 Range-edge populations have a higher relatedness and a smaller effective population size  
568 than those from central areas at ramet, and partially, at genet level. Of course, these  
569 parameters are not independent and all contribute to low genetic connectivity between edge  
570 populations and indicate the fact that trees in range-edge populations are more closely  
571 related and fewer genotypes contribute to the next generation in range-edge population than  
572 in central populations. This could be because there is less turnover in range-edge  
573 populations; genets may be very old and contributed proportionally more to the standing crop  
574 than younger trees, especially if there is a lack of sexual reproduction. In central populations  
575 there may be more turnover and more even aged genets, with more equal contribution to the  
576 next generation. Also, population fragmentation is likely to be higher at the range-edge, also  
577 lowering  $N_e$  (Chybicki *et al.*, 2014).

578            The average distance between sampled trees was not identical in all populations and  
579 could potentially bias outcomes if, for example, pairwise distances between trees from range-

580 edge populations were smaller than those from central populations. However, Av Dist only  
581 significantly affected relatedness ( $r$  ML-Relate), and only at the ramet level: this effect is as  
582 expected as the average distance between first degree related trees is 52m. Since average  
583 distance of trees is not significant as a covariate in other measures of clonal reproduction,  
584 diversity and relatedness, we are confident that the main-effect differences between range-  
585 edge and central populations are not biased due to sampling scenarios differing between  
586 sites. Moreover, the number of ramet pairs within 26m (excluding the top-quintile for clonal  
587 pairs) across all populations was similar in range-edge and in central populations in both  
588 species (182 vs 165 in *T. platyphyllos* and 594 vs 395 in *T. cordata*). The number of genet  
589 pairs within 61m (excluding the top-quintile for first degree related pairs) was also similar in  
590 range edge and in central populations (272 vs 310 in *T. platyphyllos* and 652 vs 674 in *T.*  
591 *cordata*). This indicates that there were ample potential candidates for clonal or closely  
592 related pairs, i.e. within 'critical distance', in both regions. The conclusion that relatedness  
593 between *Tilia* trees is unlikely over distances of more than 61m (top quintile), has important  
594 consequences for woodland managers.

595

#### 596 4.5 Population substructure

597 Several studies in forest trees have shown that clones or related trees can often only be  
598 expected at relatively short distances, e.g. only up to 2m in *Quercus crispula* (Aizawa *et al.*,  
599 2017). In *Tilia*, the average distance of full-sibs and parent-offspring related pairs is 52m  
600 (median 21m) and the average distance between clones is 16.8m (median 15m), which is  
601 similar to the spatial scales revealed in *Fagus grandifolia* and *Prunus avium* (Kitamura *et al.*,  
602 2003; Vaughan *et al.*, 2007) and slightly more than in tanoak (Dodd *et al.*, 2014). This means  
603 that close relatedness is only expected to be observed at a rather small scale and in  
604 populations with high density of adult trees (Duminil *et al.*, 2016). Indeed this is reflected in  
605 some populations with large distances between samples, such as NOSO, not showing any  
606 small scale population structure (data not presented).

607

#### 608 4.6 Conclusion

609 We can now attempt to answer the questions whether leading range-edge populations are at  
610 risk and whether they are suitable for range expansion. Leading range-edge populations of  
611 *Tilia* do not have a lower genetic diversity than central populations, making it likely that  
612 range-edge populations are remnants of a genetically diverse moving front, with no  
613 prolonged bottleneck effects. Although range-edge populations are more clonal than central  
614 populations, they contain medium to high genetic diversity (perhaps from an earlier period in  
615 time, e.g. Ennos, 2003) and show no inbreeding ( $F_{IS}$ ) so should be able to regenerate,  
616 naturally or assisted (Macaya-Sanz *et al.*, 2016). With the climate warming at range-edge  
617 locations sexual reproduction may become more prevalent and therefore should promote *in*  
618 *situ* adaptation to novel conditions. In addition, current populations may spread: with more  
619 continuous populations natural gene flow from more southern populations may occur.  
620 However, the effective population size is smaller at range-edge, meaning that if the  
621 fragmentation and demographics stay the same, there is a risk that genetic diversity may be  
622 lost as few individuals contribute to the next generation, while in central range populations  
623 this risk is limited as there is a more even spread of sexual reproduction amongst adult trees.  
624 Range-edge populations generally have  $N_e < 50$ , which is considered low and makes the  
625 species vulnerable due to low ability to adapt to changes (Myking, 2002), and this is  
626 exacerbated by clonal reproduction, lowering  $N_e$  to below 10. For example, the *T.*  
627 *platyphyllos* population UKAS, has small  $N_e$  and a high rate of clonal reproduction. Those  
628 types of populations seem particularly at risk, and maybe at a tipping point, due to the  
629 combined effects of strong fragmentation, human activity, and the small number of,  
630 sometimes very old, trees, belonging to few clones. Regeneration here may require good  
631 management to allow adaptation, e.g. to climate change.

632           Woodland managers and foresters wishing to sample trees for planting further north  
633 (assisted migration) can use populations that are currently at the northern range-edge of the



634 species because in those populations diversity is not lower than in central regions.  
635 Furthermore, northern populations are likely to be better adapted to conditions in the north  
636 than those from central regions, e.g. to climatic, season length and day light differences  
637 (Lobo *et al.*, 2018a). However, further work is required to test whether *Tilia* set seed at these  
638 northern locations. An alternative option for replication would be through clonal propagation,  
639 natural or assisted. The additional use of other sources, e.g. from central ranges with diverse  
640 microclimates, may generate the optimal genetic composition and diversity for adaptation. In  
641 the wake of a warming climate and a shifting natural range, successful sexual regeneration  
642 will greatly contribute to adaptation to local and novel environmental conditions (Thomas *et*  
643 *al.*, 2014).

644 Forest managers need to take into account sufficient distance between donor trees  
645 (be it seed or clonal material), *i.e.* 26m between sampling to avoid two identical genotypes,  
646 and 61m to avoid two trees that are related to the first degree (avoiding all but top quintile).  
647 Adding diverse tree species to forests has become popular, and *Tilia* in particular has shown  
648 to increase the herb layer (Normann *et al.*, 2016) and the decomposition of leaf litter (Muys *et*  
649 *al.*, 1992). Furthermore, *Tilia* is known to be a strong tree with specific associated  
650 communities performing important ecosystem services (Hommel *et al.*, 2007).

651 Here we have compared northern range-edge with central populations, but we  
652 recognise that a similar study should be performed on 'trailing edge' or 'stable rear edge' *Tilia*  
653 populations to ensure conservation of the diversity in these species (Hampe & Petit, 2005).  
654 Pinpointing the existence of such populations and obtaining samples is particularly difficult.  
655 Nonetheless, our findings may also be of importance for trailing edge populations because  
656 *Tilia* used to be one of the dominant tree species in Europe and current populations are  
657 remnants of these woodlands (e.g. UK and Denmark), and are, similarly to leading edge  
658 populations, now fragmented and limited due to human interference and climate change.

659

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673

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964  
965 **Data Accessibility Statement:** Sampling locations and microsatellite genotypes have been  
966 made available through Mendeley DOI: doi:10.17632/t7j48wg28z.1

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969 **Author contributions:**

970 SL, PP and KW collected the genotype data, SL and KW designed the experiment, collected  
971 most of the samples, performed the genetic data analyses and wrote the manuscript, RS  
972 performed the GLM statistical analyses. All authors read and commented on the ms.

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976 **Table 1** *Tilia* populations, ordered by location codes within groups, country, average distance  
 977 (Av Dist) between trees in m, number (N) and coordinates of trees: non-shaded blocks are  
 978 leading range-edge populations, while shaded blocks are central range populations (na not  
 979 assessed)

Species/Code	Location	Country	Av Dist	N	Latitude (°N)	Longitude (°E)
<i>T. platyphyllos</i>						
DEBOp	Bolderslev	Denmark	38	11	54.9978	9.3512
SWWC	West Coast	Sweden	na	15	58.8967	11.0419
UKAS	Aplegarth	UK	67	13	54.4093	-1.8165
UKAW	Anston Wd	UK	54	15	53.3402	-1.1984
UKHD	Hudswell	UK	269	25	54.4012	-1.7573
UKHW	Halesend Wd	UK	52	24	52.1407	-2.3815
AUDOp	Dobra	Austria	27	14	48.5907	15.3974
AULE	Leopoldsberg	Austria	91	25	48.2774	16.3522
AUSOp	Sommerein	Austria	58	14	47.9848	16.6947
FRML	Mont Lure	France	1250	20	44.1252	5.8705
FRVE	Ventoux	France	374	20	44.1869	5.2382
GELC	Lichtenstein	Germany	362	31	48.4071	9.2627
<i>T. cordata</i>						
DEBOc	Bolderslev	Denmark	55	30	54.9978	9.3516
FIIS	Iso Kirvessaari	Finland	49	20	62.5548	27.7626
FINI	Niinisaar	Finland	36	20	61.8171	29.3895
NOSO	Sogn Fjordane	Norway	854	16	61.8505	6.1375
RUVA	Vagay area	Russia	na	20	57.5096	69.1954
SWOL	Øland	Sweden	71	27	56.6070	16.4503
UKBB	Brignal Banks	UK	36	19	54.4968	-1.9114
UKRO	Roudsea	UK	47	40	54.2337	-3.0260
AUDOc	Dobra	Austria	50	21	48.5909	15.3975
AUSOc	Sommerein	Austria	83	23	47.9848	16.6947
AUST	Stams	Austria	106	21	47.2757	10.9772
AUTH	Thayatal	Austria	725	17	48.8448	15.8878
CZVO	Velky Osek	Czech Repub	67	20	50.1012	15.1788
FRMO	Mouthiers	France	69	20	48.9120	4.9138
GECO	Colbitz	Germany	270	20	52.3310	11.5559
POBF	Białowieża	Poland	930	40	52.7272	23.8351

980

981 **Table 2** Means of estimates of clonal occurrence in *Tilia* and significance of fixed and  
 982 covariate effects.

Species/region/ <i>P</i> -values	<i>N</i>	<i>G</i>	<i>R</i>	<i>D</i> *
<i>T. platyphyllos</i>				
Range-edge	17.2	14.7	0.804	0.947
Central	21.0	20.7	0.981	0.998
<i>T. cordata</i>				
Range-edge	24.0	17.6	0.685	0.937
Central	23.0	23.0	1.000	1.000
<i>P</i> -values				
Region			<b>&lt; 0.0005</b>	<b>&lt; 0.0005</b>
Species			0.143	0.186
Av Dist			0.053	0.099
Region x species interaction			0.291	0.425

983 **Footnote:** Number of samples (*N*), number of genotypes (*G*) and clonal occurrence  
 984 presented as genotypic richness  $R = (G-1)/(N-1)$  and Simpson's complement index for  
 985 genotypic diversity *D*\*, from range-edge and central locations, *P* values for significance of  
 986 fixed-effect of region and species, co-variate effect of the average distance between samples  
 987 (Av Dist), interaction of species and region effect, significant effects are indicated in bold  
 988  
 989

990 **Table 3** Genetic characteristics of *Tilia* as means per group of populations at the genet level  
 991 and outcomes of statistical tests (*P* values)

Species/region	$H_e$	$F_{IS}$
<i>T. platyphyllos</i>		
Range-edge	0.690	-0.061
Central	0.745	-0.024
<i>T. cordata</i>		
Range-edge	0.553	-0.037
Central	0.573	0.013
<i>P</i> -values		
Effect region	0.319	0.545
Effect species	<b>&lt; 0.0005</b>	0.729
Av Dist	0.373	NA
Region x species interaction	0.161	0.874

992 **Footnote:** Expected heterozygosity ( $H_e$ ) and fixation index ( $F_{IS}$ ) and *P*-values for fixed effects  
 993 of region, species and interaction effects, as well as significance of the average distance  
 994 between samples (Av Dist) within a population, used as a covariate, significant effects are  
 995 indicated in bold.  
 996

**Table 4.** Expected heterozygosity  $H_e$  in all genotypes of a population, observed heterozygosity  $H_o$  and fixation index  $F_{IS}$  of non-clonal and clonal MLL in six populations of *T. cordata*, with number of samples in a group indicated as n.

Population	$H_e$ over all (n)	$H_o$ non-clonal (n)	$H_o$ clonal (n)	$F_{IS}$ non-clonal genets	$F_{IS}$ clonal genets
DEBOc	0.575 (18)	0.545 (13)	0.550 (5)	0.052	0.043
FIIS	0.505 (12)	0.552 (8)	0.583 (4)	-0.093	-0.155
FINI	0.499 (13)	0.481 (9)	0.583 (4)	0.035	-0.169
RUVA	0.576 (14)	0.483 (10)	0.563 (4)	0.161	0.024
UKBB	0.520 (6)	0.556 (3)	0.639 (3)	-0.069	-0.229
UKRO	0.566 (29)	0.561 (19)	0.600 (10)	0.008	-0.060
<b>Average</b>	<b>0.540</b>	<b>0.530</b>	<b>0.586</b>	<b>0.016</b>	<b>-0.091</b>

**Table 5.** Diversity and G-statistics of leading range-edge and central populations of *Tilia* species and *P*-values for comparisons between the regions (GenoDive 2.0b) with 95% confidence intervals (bootstrapping) between brackets at the genet level, significant effects are indicated in bold

Species/region	$H_o$	$H_s$	$G_{IS}$	$G_{ST}$	$G'_{ST}$	$D_{est}$	Clonal div
<i>T. platyphyllos</i>							
Range-edge	0.732 (0.651-0.805)	0.72 (0.652-0.772)	-0.017 (-0.064-0.032)	0.108 (0.092-0.124)	0.442 (0.325-0.564)	0.375 (0.252-0.507)	0.951
Central	0.764 (0.649-0.861)	0.760 (0.648-0.850)	-0.005 (-0.042-0.034)	0.049 (0.041-0.060)	0.234 (0.166-0.350)	0.195 (0.123-0.320)	0.998
<i>P</i> -value	0.054	<b>0.035</b>	0.657	<b>0.014</b>	<b>0.013</b>	<b>0.013</b>	0.068
<i>T. cordata</i>							
Range-edge	0.570 (0.420-0.699)	0.575 (0.424-0.705)	0.008 (-0.050-0.069)	0.084 (0.064-0.104)	0.214 (0.135-0.331)	0.142 (0.070-0.261)	0.946
Central	0.558 (0.406-0.694)	0.587 (0.425-0.731)	0.050 (-0.015-0.126)	0.048 (0.030-0.070)	0.126 (0.069-0.206)	0.082 (0.036-0.158)	1
<i>P</i> -value	0.644	0.525	0.189	0.064	<b>0.050</b>	<b>0.040</b>	<b>0.003</b>

$H_o$  observed heterozygosity and  $H_s$  expected heterozygosity within populations,  $G_{IS}$  inbreeding coefficient and  $G_{ST}$  fixation index,  $G'_{ST}$  is the fixation index corrected for bias due to limited number of populations, and Jost's  $D_{est}$  population differentiation independent from  $H_s$ , and clonal diversity as measure of amount of sexual reproduction, analogous to Simpsons  $D^*$

**Table 6.** Distribution of genetic diversity (AMOVA) within and among groups of populations in *Tilia* species at the genet level (GenoDive 2.0b). Standard deviations (St dev) of F-statistics were obtained through jack-knifing over loci and 95% confidence intervals of F-statistics were obtained through bootstrapping over loci. P values indicate significance of F-value > 0.

	%var	F-statistic	F-value	St dev	P
<i>T. platyphyllos</i>					
<b>Range edge</b>					
Within individuals	0.886	$F_{it}$	0.114 (0.063-0.167)	0.028	-
Among individuals	0.002	$F_{is}$	0.002 (-0.050-0.059)	0.029	0.486
Among populations	0.112	$F_{ST}$	0.112 (0.098-0.127)	0.008	0.001
<b>Central</b>					
Within individuals	0.940	$F_{it}$	0.053 (0.016-0.096)	0.210	-
Among individuals	0.003	$F_{is}$	0.003 (-0.038-0.034)	0.019	0.631
Among populations	0.055	$F_{ST}$	0.055 (0.046-0.068)	0.006	0.001
<i>T. cordata</i>					
<b>Range edge</b>					
Within individuals	0.900	$F_{it}$	0.099 (0.044-0.148)	0.027	-
Among individuals	0.008	$F_{is}$	0.099 (-0.048-0.065)	0.031	0.306
Among population	0.091	$F_{ST}$	0.091 (0.066-0.115)	0.013	0.001
<b>Central</b>					
Within individuals	0.903	$F_{it}$	0.097 (0.027-0.176)	0.041	-
Among individuals	0.041	$F_{is}$	0.043 (-0.022-0.117)	0.038	0.001
Among population	0.056	$F_{ST}$	0.056 (0.038-0.079)	0.011	0.001

**Table 7** Means or medians of relatedness characteristics of *Tilia*, at the genet level and outcomes of statistical tests for region, species and interaction effects (*P* values, significant effects are indicated in bold).

	<i>r</i> ML-Relate mean	1 <sup>st</sup> +2 <sup>nd</sup> degr rel mean	1 <sup>st</sup> degr rel mean	<i>N<sub>e</sub></i> Co-an median	<i>N<sub>e</sub></i> LD median
<i>T. platyphyllos</i>					
Range-edge	0.060	0.060	0.027	8.7	16.2
Central	0.038	0.042	0.013	48.1	44.5
<i>T. cordata</i>					
Range-edge	0.054	0.060	0.023	60.9	38.2
Central	0.046	0.048	0.009	∞	494.5
<i>P</i> -values					
Effect region	0.236	0.120	<b>0.003</b>	<b>0.049</b>	<b>0.031</b>
Effect species	0.079	0.269	0.674	<b>0.009</b>	0.078
Av. Dist.	0.381	0.697	0.518	0.578	0.889
Region x species interaction	0.134	0.215	0.180	0.885	0.292

**Footnote:** Shown are the average relatedness of individuals (*r*, ML-Relate), proportion of first and second degree related individuals, proportion of first degree related individuals and effective population size (*N<sub>e</sub>*) calculated based on Co-ancestry and on Linkage Disequilibrium (LD).

**Table 8** Means or medians of relatedness characteristics of *Tilia*, at the ramet (sample) level and outcomes of statistical tests for region, species and interaction effects (*P* values, significant effects are indicated in bold).

	<i>r</i> ML- Relate mean	1 <sup>st</sup> +2 <sup>nd</sup> degr rel mean	1 <sup>st</sup> degr rel mean	<i>N<sub>e</sub></i> Co-an median	<i>N<sub>e</sub></i> LD median
<i>T. platyphyllos</i>					
Range-edge	0.109	0.060	0.027	9.4	5.85
Central	0.040	0.042	0.013	48.2	37.1
<i>T. cordata</i>					
Range-edge	0.108	0.060	0.023	8.8	3.0
Central	0.046	0.048	0.009	∞	494.5
<i>P</i> -values					
Effect region	<b>&lt;0.0005</b>	0.519	<b>0.045</b>	<b>0.005</b>	<b>&lt; 0.0005</b>
Effect species	0.279	0.154	0.453	<b>0.048</b>	<b>0.042</b>
Av. Dist.	<b>0.040</b>	0.942	0.474	0.335	0.352
Region x species interaction	0.688	0.349	0.132	0.425	0.079

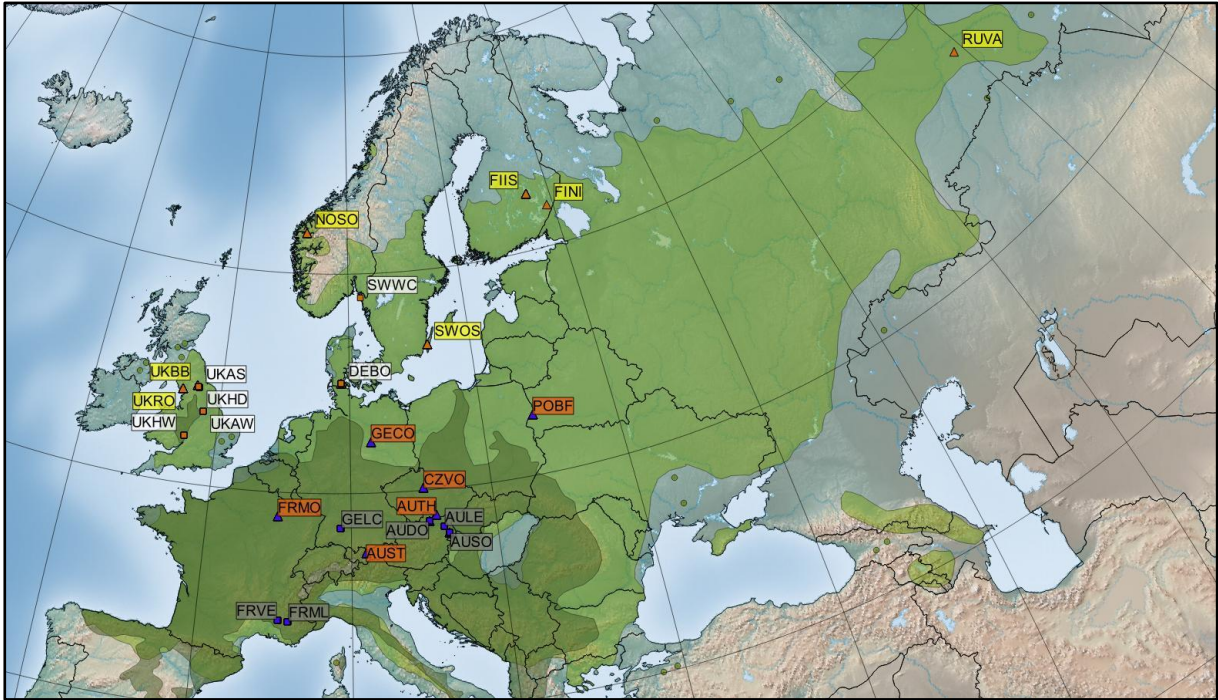
**Footnote:** Shown are the average relatedness of individuals (*r*, ML-Relate), proportion of first and second degree related individuals, proportion of first degree related individuals and effective population size (*N<sub>e</sub>*) calculated based on Co-ancestry and on Linkage Disequilibrium (LD)



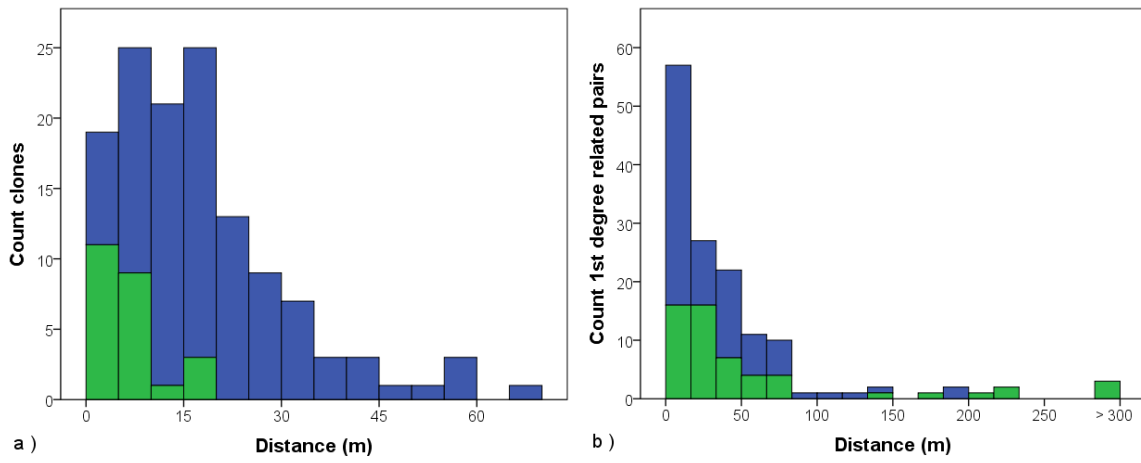
Figure legends:

**Fig 1.** Distribution of *Tilia cordata* (light green area and dots) and *T. platyphyllos* (dark green area and dots). Distribution data from EUFORGEN ([www.euforgen.org](http://www.euforgen.org)). Map constructed using QGIS v2.16.2 ([www.qgis.org](http://www.qgis.org)). Triangles are *T. cordata* populations sampled while circles are *T. platyphyllos*. Orange icons show leading range-edge populations, for *T. cordata* (>54°N) with yellow background and for *T. platyphyllos* (>52°N) with white background. Blue icons are populations from central range of the two species (>43°N), *T. cordata* with orange background and *T. platyphyllos* with grey background.

**Figure 2** Counts of numbers of a) clone pairs, in *T. platyphyllos* (green) and *T. cordata* (blue) across distance classes b) first degree related pairs of individuals (full sib and parent-offspring), three pairs with distance of 483, 684 and 809m are combined into >300m class



**Fig 1.** Distribution of *Tilia cordata* (light green area and dots) and *T. platyphyllos* (dark green area and dots). Distribution data from EUFORGEN ([www.euforgen.org](http://www.euforgen.org)). Map constructed using QGIS v2.16.2 ([www.qgis.org](http://www.qgis.org)). Triangles are *T. cordata* populations sampled while circles are *T. platyphyllos*. Orange icons show leading range-edge populations, for *T. cordata* (>54°N) with yellow background and for *T. platyphyllos* (>52°N) with white background. Blue icons are populations from central range of the two species (>43°N), *T. cordata* with orange background and *T. platyphyllos* with grey background.



**Figure 2** Counts of numbers of a) clone pairs b) first degree related pairs of individuals (full sib and parent-offspring), in *T. platyphyllos* (green) and *T. cordata* (blue) across distance classes. Three related pairs with distances of 483, 684 and 809m are combined into the >300m class