



# Whole-Genome Sequence and Fosfomycin Resistance of *Bacillus* sp. Strain G3(2015) Isolated from Seawater off the Coast of Malaysia

Xin-Yue Chan,<sup>a</sup> Jian-Woon Chen,<sup>a,b</sup> Tan-Guan-Sheng Adrian,<sup>a</sup> Kar-Wai Hong,<sup>a</sup> Chien-Yi Chang,<sup>c,d</sup> Wai-Fong Yin,<sup>a</sup>  Kok-Gan Chan<sup>a,b</sup>

Division of Genetics and Molecular Biology, Institute of Biological Sciences, Faculty of Science, University of Malaya, Kuala Lumpur, Malaysia<sup>a</sup>; UM Omics Centre, University of Malaya, Kuala Lumpur, Malaysia<sup>b</sup>; Institute of Life and Earth Sciences, Heriot Watt University, Edinburgh, United Kingdom<sup>c</sup>; Guangzhou Institute of Pediatrics, Guangzhou Women and Children's Medical Center, Guangzhou Medical University, Guangzhou, Guangdong, China<sup>d</sup>

**ABSTRACT** *Bacillus* sp. is a Gram-positive bacterium that is commonly found in seawater. In this study, the genome of marine *Bacillus* sp. strain G3(2015) was sequenced using MiSeq. The fosfomycin resistant gene *fosB* was identified upon bacterial genome annotation.

The high adaptability of *Bacillus* spp. enables their survival in a wide range of environments, including environments with extreme temperature, pressure, salinity, and pH (1). The production of diverse classes of antibiotics by *Bacillus* spp. has enhanced their ability to compete for survival in diverse environments (2, 3). In contrast to its extensively studied terrestrial counterparts, less research has been conducted on marine *Bacillus* spp. In this study, we present the whole genome of marine *Bacillus* sp. strain G3(2015) and its antibiotic resistance gene.

*Bacillus* sp. G3(2015) was isolated from subsurface seawater collected from the coast of Georgetown, Malaysia. Genomic DNA was extracted from *Bacillus* sp. G3(2015) using the MasterPure Gram-positive DNA purification kit (Epicenter, USA). The extracted DNA was quantified using a Qubit 2.0 Fluorometer (Thermo Fisher Scientific, USA) while the quality of the DNA was verified using a NanoDrop spectrophotometer (Thermo Fisher Scientific, USA) (4–6). Sequencing library preparation was prepared using a Nextera DNA library preparation kit (Illumina, USA). Whole-genome sequencing was performed on the MiSeq sequencer (Illumina, USA) using MiSeq reagent kit v2. Paired-end reads generated from MiSeq were trimmed and *de novo* assembled using CLC Genomic Workbench (v8). Gene prediction was carried out using Prodigal (v2.6.1) (7). The bacterial genome was then annotated using the NCBI Prokaryotic Genome Annotation Pipeline (v3) and Rapid Annotations using Subsystems Technology (RAST) (v2) (8, 9). The predicted gene sequences were also aligned using BLAST against the Antibiotic Resistance Genes Database (ARDB) to determine the presence of antibiotic resistance genes (10).

The bacterial genome was assembled into 120 contigs with an average coverage of 75-fold. The  $N_{50}$  of the contigs is 83,170 bp. The estimated genome size is 5.4 Mbp with 34.9% G+C content. A total of 5,565 coding sequences (CDS), eight rRNAs, 96 tRNAs, and 99 pseudogenes were identified from the draft genome.

Fosfomycin resistance gene (*fosB*) (accession number WP\_000943769) was detected in contig 76 of the genome (NZ\_LKID01000076.1) of *Bacillus* strain G3(2015) by both RAST and BLAST against ARDB. Fosfomycin is an antibiotic functionally effective against Gram-positive and Gram-negative bacteria by irreversibly inhibiting bacterial cell wall

Received 21 January 2017 Accepted 24 January 2017 Published 30 March 2017

**Citation** Chan X-Y, Chen J-W, Adrian T-G-S, Hong K-W, Chang C-Y, Yin W-F, Chan K-G. 2017. Whole-genome sequence and fosfomycin resistance of *Bacillus* sp. strain G3(2015) isolated from seawater off the coast of Malaysia. Genome Announc 5:e00067-17. <https://doi.org/10.1128/genomeA.00067-17>.

**Copyright** © 2017 Chan et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Kok-Gan Chan, [kokgan@um.edu.my](mailto:kokgan@um.edu.my).

biosynthesis (11, 12). It is mainly prescribed for the treatment of uncomplicated urinary tract infections and gastrointestinal infections (12). The *fosB* gene synthesizes thiol-S-transferase (FosB) which catalyzes the addition of bacillithiol to the fosfomycin epoxide ring leading to the inhibition of fosfomycin (13). Studies have shown that the *fosB* gene is transferable across genera via conjugation (14, 15). The high bacterial diversity in tropical seawater could increase the rate of emergence of fosfomycin resistant strains from different genera. Therefore, this study on fosfomycin resistant *Bacillus* sp. strain G3(2015) that was isolated from seawater is important, especially in the context of public health.

**Accession number(s).** This whole-genome shotgun project has been deposited in DDBJ/EMBL/GenBank under the accession no. [LKID00000000](https://doi.org/10.1128/MMBR.64.3.548-572.2000). The version described in this paper is the first version, LKID01000000.

## ACKNOWLEDGMENTS

This study was supported by research grants from University of Malaya through HIR grants awarded to K.-G.C. (UM-MOHE HIR grant UM C/625/1/HIR/MOHE/CHAN/14/1, H-50001-A000027; UM-MOHE HIR grant UM C/625/1/HIR/MOHE/CHAN/01, A000001-50001) and Postgraduate Research grant PG083-2015B which was awarded to X.-Y.C.

## REFERENCES

- Nicholson WL, Munakata N, Horneck G, Melosh HJ, Setlow P. 2000. Resistance of *Bacillus* endospores to extreme terrestrial and extraterrestrial environments. *Microbiol Mol Biol Rev* 64:548–572. <https://doi.org/10.1128/MMBR.64.3.548-572.2000>.
- Cao M, Helmann JD. 2002. Regulation of the *Bacillus subtilis* bcrC bacitracin resistance gene by two extracytoplasmic function  $\sigma$  factors. *J Bacteriol* 184:6123–6129. <https://doi.org/10.1128/JB.184.22.6123-6129.2002>.
- Adimpong DB, Sørensen KI, Thorsen L, Stuer-Lauridsen B, Abdelgadir WS, Nielsen DS, Derckx PMF, Jespersen L. 2012. Antimicrobial susceptibility of *Bacillus* strains isolated from primary starters for African traditional bread production and characterization of the bacitracin operon and bacitracin biosynthesis. *Appl Environ Microbiol* 78:7903–7914. <https://doi.org/10.1128/AEM.00730-12>.
- Chan XY, Chua KH, Yin WF, Puthuchery SD, Chan KG. 2014. Whole-genome analysis of *Aeromonas hydrophila* strain 187, exhibiting quorum-sensing activity. *Genome Announc* 2(6):e01360-14. <https://doi.org/10.1128/genomeA.01360-14>.
- Chen JW, Gan HM, Yin WF, Chan KG. 2012. Genome sequence of *Roseomonas* sp. strain B5, a quorum-quenching *N*-acylhomoserine lactone-degrading bacterium isolated from Malaysian tropical soil. *J Bacteriol* 194:6681–6682. <https://doi.org/10.1128/JB.01866-12>.
- Hong KW, Gan HM, Low SM, Lee PKY, Chong YM, Yin WF, Chan KG. 2012. Draft genome sequence of *Pantoea* sp. strain A4, a rafflesia-associated bacterium that produces *N*-acylhomoserine lactones as quorum-sensing molecules. *J Bacteriol* 194:6610. <https://doi.org/10.1128/JB.01619-12>.
- Hyatt D, Chen GL, LoCascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC Bioinformatics* 11:119. <https://doi.org/10.1186/1471-2105-11-119>.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Ciufu S, Li W. 2013. Prokaryotic genome annotation pipeline. *In* The NCBI Handbook, 2nd ed. National Center for Biotechnology Information, Bethesda, MD. <http://www.ncbi.nlm.nih.gov/books/NBK174280>.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). *Nucleic Acids Res* 42:D206–D214. <https://doi.org/10.1093/nar/gkt1226>.
- Liu B, Pop M. 2009. ARDB-antibiotic resistance genes database. *Nucleic Acids Res* 37:D443–D447. <https://doi.org/10.1093/nar/gkn656>.
- Mascher T, Margulis NG, Wang T, Ye RW, Helmann JD. 2003. Cell wall stress responses in *Bacillus subtilis*: the regulatory network of the bacitracin stimulon. *Mol Microbiol* 50:1591–1604. <https://doi.org/10.1046/j.1365-2958.2003.03786.x>.
- Thompson MK, Keithly ME, Harp J, Cook PD, Jagessar KL, Sulikowski GA, Armstrong RN. 2013. Structural and chemical aspects of resistance to the antibiotic fosfomycin conferred by FosB from *Bacillus cereus*. *Biochemistry* 52:7350–7362. <https://doi.org/10.1021/bi4009648>.
- Roberts AA, Sharma SV, Strankman AW, Duran SR, Rawat M, Hamilton CJ. 2013. Mechanistic studies of FosB: a divalent-metal-dependent bacillithiol-S-transferase that mediates fosfomycin resistance in *Staphylococcus aureus*. *Biochem J* 451:69–79. <https://doi.org/10.1042/BJ20121541>.
- Xu X, Chen C, Lin D, Guo Q, Hu F, Zhu D, Li G, Wang M. 2013. The fosfomycin resistance gene *fosB3* is located on a transferable, extrachromosomal circular intermediate in clinical *Enterococcus faecium* isolates. *PLoS One* 8:e78106. <https://doi.org/10.1371/journal.pone.0078106>.
- Suárez JE, Mendoza MC. 1991. Plasmid-encoded fosfomycin resistance. *Antimicrob Agents Chemother* 35:791–795. <https://doi.org/10.1128/AAC.35.5.791>.